Related ELISA kits available from ADI (see details at the web site)

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**Anti-Bordetella pertussis IgG ELISA Kit**

**Cat. # 960-110-PHG**

For the detection of Anti-Bordetella pertussis IgG In Serum/Plasma

For In Vitro Research Use Only

**Instruction Manual No. M-960-110-PHG**

**ALPHA DIAGNOSTIC INTERNATIONAL**

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Toll Free (800) 786-5777

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Web Site: www.4adi.com

Related ELISA kits available from ADI (see details at the web site)
Anti-Bordetella pertussis IgG ELISA KIT #960-110-PHG

This kit has been designed for the detection of Anti-Bordetella pertussis IgG serum or plasma. For research use only, not for use in diagnostic procedures.

<table>
<thead>
<tr>
<th>Kit Components</th>
<th>Cat. #</th>
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<tbody>
<tr>
<td>B. pertussis antigens coated ELISA strips (96 wells)</td>
<td>960111</td>
</tr>
<tr>
<td>Calibrator A, Negative Control (1 U/ml), 2 ml</td>
<td>960112A</td>
</tr>
<tr>
<td>Calibrator B, Cut-off Standard (20 U/ml), 2 ml</td>
<td>960112B</td>
</tr>
<tr>
<td>Calibrator C, Weak-positive Control (45 U/ml), 2 ml</td>
<td>960112C</td>
</tr>
<tr>
<td>Calibrator D, Positive Control, (150 U/ml), 2 ml</td>
<td>960112D</td>
</tr>
</tbody>
</table>

Controls are diluted serum base containing 0.01% BND as preservative

Sample Diluent, 60 ml buffer with 0.09% azide 960110-SD
Wash buffer (10X), 60 ml 960110-WB
Anti-hlgG HRP Conjugate, 15 ml, ready to use 960113
HRP Substrate Soln (TMB), 15 ml, ready to use 960110-TM
Stop Solution (diluted sulfuric acid), 15 ml 960110-SS
Complete Instruction Manual M-960110PHG

General Information:

Whooping cough is a disease of the respiratory tracts which is caused by Bordetella pertussis bacteria. It is transmitted by airborne infection. The gramnegative Cocccobacillus produces a series of biologically active molecules. The different compounds appear either during the pathogenesis or during the process of immunization against pertussis and show different effects. A characterisation has been made for the pertussis toxin (PT), the filamentery haemagglutinine (FHA) and different lipopolysaccharides (LPS). Pertussis shows a high rate of transmission (rates of infection of over 90 % have been found for nonvaccinated household members) and can cause severe diseases, especially for very young children. From 10749 patients under one year between 1980 and 1989 69 % were brought into hospital, 22 % suffered from pneumonia, 0.9 % showed an Encephalopathy and 0.6 % died. For older children and adults (including already vaccinated persons) the infection may be observed by an unspecified bronchitis or inflammation of the upper respiratory tracts. Even asymptomatic cases are quite common.

The serological response following pertussis disease or immunization with pertussis vaccine has been measured with agglutination assays, precipitins, complement fixation and enzyme-linked immunosorbent assay (ELISA). Enzyme-linked immunosorbent assays, in which Bordetella antigen (containing toxin, FHA and LPS and standardized in U/ml) is bound to a solid phase support, are sensitive, easy to perform and can be used both to determine seropositivity with a single serum and to indicate recent Bordetella infection by determination of IgM and IgA.

Pertussis Vaccines: Trihibit (DTAP/Hib), ActHib (Hib-PRP-T), Daptec (DTAP), Tripedia (DTAP), Adacel (tetanus, Diphtheria, Acellular Pertussis) - Sanofi Pasteur; PedvaxHib (Hib-PRP-OMP) – Merck; Pediarix (DTAP/HepB/IPV), Infanrix (DTAP), Boostrix (Tetanus, Diphtheria, Acellular Pertussis) - GlaxoSmithKline.

Interpretation of results:

Most of the data presented here is for information purpose. Therefore, users are suggested to establish their own reference values.

<table>
<thead>
<tr>
<th>U/mL</th>
<th>Interpretation</th>
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<tbody>
<tr>
<td>&lt; 18</td>
<td>negative</td>
</tr>
<tr>
<td>18 - 22</td>
<td>equivocal</td>
</tr>
<tr>
<td>&gt; 22</td>
<td>positive</td>
</tr>
</tbody>
</table>

The results themselves should not be the only reason for any therapeutical consequences. They have to be correlated to other clinical observations and diagnostic tests.

Expected Values:

In an in-house study apparently healthy subjects showed the following results:

<table>
<thead>
<tr>
<th>Ig Isotype</th>
<th>n</th>
<th>Interpretation</th>
</tr>
</thead>
<tbody>
<tr>
<td>IgG</td>
<td>54</td>
<td>positive</td>
</tr>
<tr>
<td></td>
<td></td>
<td>equivocal</td>
</tr>
<tr>
<td></td>
<td></td>
<td>Negative</td>
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</tbody>
</table>

PERFORMANCE CHARACTERISTICS

Bordetella pertussis ELISA IgG

Intra-Assay-Precision 5.0 %
Inter-Assay-Precision 4.3 %
Inter-Lot-Precision 2.6 – 4.5 %
Analytical Sensitivity 0.98 U/mL
Recovery 106 – 114 %
Linearity 78 – 124 %

Cross-Reactivity: No cross-reactivity to RSV, Adenovirus and Parainfluenza IgG

Interferences: No interferences to bilirubin up to 0.3 mg/mL, hemoglobin up to 8.0 mg/mL und triglycerides up to 5.0 mg/mL

Clinical Specificity 84 %
Clinical Sensitivity 100 %

Specificity of the Antigens

The coated antigens in this kit contain B. Pertussis antigens. It has B. pertussis toxin, filamentous hemeagglutinin (FHA), pertactin (PRN), LPS among others. Therefore this kit will detect antibodies to all major B. pertussis antigens. ADI has other kits that has purified Pertussis toxin or FHA or PRN antigens to investigate the presence of specific antibodies (see the list on page 8).

Species Reactivity

This kit is designed for human serum or plasma. ADI also has kits for mouse, rabbit and other species.


Alpha Diagnostic Intl. (www.4adi.com) 960-110-PHG/130828A
**WORKSHEET OF TYPICAL ASSAY**

<table>
<thead>
<tr>
<th>Wells</th>
<th>Stds/samples (U/ml)</th>
<th>Net Mean A\textsubscript{450 nm}</th>
<th>Calculated Conc. (U/ml)</th>
</tr>
</thead>
<tbody>
<tr>
<td>A1, A2</td>
<td>Std. A negative control (1 U/ml)</td>
<td>0.042</td>
<td></td>
</tr>
<tr>
<td>B1, B2</td>
<td>Std. B Cut-off (20 U/ml)</td>
<td>0.483</td>
<td></td>
</tr>
<tr>
<td>C1, C2</td>
<td>Std. C Weak positive control (45 U/ml)</td>
<td>0.786</td>
<td></td>
</tr>
<tr>
<td>D1, D2</td>
<td>Std. D positive control (150 U/ml)</td>
<td>1.789</td>
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</table>

**NOTE:** These data are for demonstration purpose only. A complete standard curve must be run in every assay to determine sample values. Each laboratory should determine their own normal reference values.

**INTERPRETATION AND CALCULATION OF RESULTS**

The obtained OD of the standards (y-axis, linear) are plotted against their concentration (x-axis, logarithmic) either on semi-logarithmic graph paper or using an automated method. A good fit is provided with cubic spline, 4 parameter logistics or Logit-Log. For the calculation of the standard curve apply each signal of the standards (one obvious outlier of duplicates might be omitted and the more plausible single value might be used). The concentration of the samples can be read from the standards curve. The initial dilution has been taken into consideration when reading the results from the graph. Results of samples of higher predilution (1:100) have to be multiplied with the dilution factor. Samples showing concentrations above the highest standard have to be diluted as described in “Assay identification of the dyed bacteria in sputum. Recently specific antigens have been prepared either by purification of natural material or by recombinant methods.

Additional ELISA kits to detect the Mycobacterium tuberculosis virus antibody in mouse and other species are also available for research. These kits should be useful to determine the M. tuberculosis antibodies due to natural infection or upon vaccination with BCG vaccine.

**PRINCIPLE OF THE TEST**

Anti-Bordetella pertussis IgG ELISA kit is based on binding of antibody from serum samples to M. tuberculosis antigens immobilized on microtiter wells. After a washing step, anti-IgG-HRP conjugate is added. After another washing step to remove all the unbound enzyme conjugate, chromogenic substrate (TMB) is added and color developed. The enzymatic reaction (color) is directly proportional to the amount of IgG present in the sample. Adding stopping solution terminates the reaction. Absorbance is then measured on a microtiter well ELISA reader at 450 nm and the concentration of IgG in samples is calculated compared with the absorbance of the supplied negative and positive controls.

**MATERIALS AND EQUIPMENT REQUIRED**

Adjustable micropipet (5-1000 μl) and multichannel pipet with disposable plastic tips. Reagent troughs, plate washer (recommended) and ELISA plate Reader.

**PRECAUTIONS**

This ELISA test is intended for in vitro research use only. The reagents contains human serum and preservative; necessary care should be taken when disposing solutions. Human sera are shown to be negative for HBsAg and HIV antibodies. Nevertheless, such tests are unable to prove the complete absence of viruses, therefore, sera should be handled with appropriate precautions.

Applicable MSDS, if not already on file, for the following reagents can be obtained from ADI or the web site.

TMB (substrate), H\textsubscript{2}SO\textsubscript{4} (stop solution), and BND (0.1% v/v in standards, sample diluent and HRP-conjugates). All waste material should be properly disinfected before disposal. Avoid contact with the stop solution (diluted sulfuric acid).

**SAMPLE COLLECTION AND HANDLING**

Blood should be collected by venipuncture, allowed clot, and serum separated by centrifugation at room temperature. Do not heat inactivate the serum. If sera can not be immediately assayed, these could be stored at -20°C for up to six months. Avoid repeated freezing and thawing of samples. No preservatives should be added to the serum. EDTA/Heparin plasma can also be used.

**Preparation of the reagent:**

Dilute wash buffer (1:10) with distilled water (60 ml stock in total of 600 ml water). Store at 4°C. If stock shows crystal then it can be dissolved by bringing to room temp or slight warming.
STORAGE AND STABILITY

The microtiter well plate and all other reagents are stable at 2-8°C until the expiration date printed on the label. The whole kit stability is usually 12 months from the date of shipping under appropriate storage conditions. Do not contaminate the bottles. Withdraw solutions in a separate clean tube or dispensing trays. Any unused solution should be discarded and not returned to the bottle. Do not use HRP substrate solution if this solution is blue. Do not expose these solutions to strong light.

TEST PROCEDURE (ALLOW ALL REAGENTS TO REACH ROOM TEMP. BEFORE USE).

1. Label, and secure the microtiter well strips to be used on the plate. Dilute samples (1:101) in sample diluent. Controls provided in the kit are already pre-diluted.

2. Pipet 100 ul of sample diluent (for use as blanks), pre-diluted negative, positive controls, and diluted serum samples into appropriate wells in duplicate. Mix gently for 5-10 seconds, cover the plate and incubate for 60 minutes at room temp (24-28oC).

3. Aspirate and wash the wells 3 times with 300 ul of diluted wash buffer. We recommend using an automated ELISA plate washer for better consistency. Failure to wash the wells properly will lead to high blank values. If washing manually, plate must be tapped over paper towel between washings to ensure proper washing.

4. Add 100 ul of antibody-enzyme conjugate into each well. Mix gently for 5-10 seconds. Cover the plate and incubate for 30 minutes at room temp.

5. Aspirate and wash the wells 3 times as above.

6. Dispense 100 ul TMB substrate per well. Mix gently for 5 seconds. Cover the plate and incubate at room temp in the dark. for 20 minutes. Blue color develops in positive wells.

7. Stop the reaction by adding 100 ul of stopping solution to all wells at the same timed intervals. Mix gently for 5-10 seconds. Blue color turns yellow. Measure the absorbance at 450 nm using an ELISA reader.

Limitations, Precautions and General Comments:

- Only for in-vitro use!
- Do not ingest or swallow! The usual laboratory safety precautions as well as the prohibition of eating, drinking and smoking in the lab have to be followed.
- All sera and plasma or buffers based upon, have been tested respective to HBsAg, HIV and HCV with recognized methods and were found negative. Nevertheless precautions like the use of latex gloves have to be taken.

- Serum and reagent spills have to be wiped off with a disinfecting solution (e.g. sodium hypochlorite, 5%) and have to be disposed of properly.

- All reagents have to be brought to room temperature (18 to 25 °C) before performing the test.

- Before pipetting all reagents should be mixed thoroughly by gentle tilting or swinging. Vigorous shaking with formation of foam should be avoided.

- It is important to pipet with constant intervals, so that all the wells of the microtiter plate have the same conditions.

- When removing reagents out of the bottles, care has to be taken that the stoppers are not contaminated. Further a possible mix-up has to be avoided. The content of the bottles is usually sensitive to oxidation, so that they should be opened only for a short time.

- In order to avoid a carry-over or a cross-contamination, separate disposable pipet tips have to be used.

- No reagents from different kit lots have to be used, they should not be mixed among one another.

- All reagents have to be used within the expiry period.

- In accordance with a Good Laboratory Practice (GLP) or following ISO9001 all laboratory devices employed should be regularly checked regarding the accuracy and precision. This refers amongst others to microliter pipets and washing or reading (ELISA-Reader) instrumentation.

- The contact of certain reagents, above all the stopping solution and the substrate with skin, eye and mucosa has to be avoided, because possible irritations and acid burns could arise, and there exists a danger of intoxication.