

## INTENDED USE

The **Human Anti-Rabies IgG** ELISA Kit detects and quantifies rabies virus -specific IgG in human serum or plasma of vaccinated, immunized and/or infected hosts. This immunoassay is suitable for:

- Determining **immune status** relative to non-immune controls
- Assessing efficacy of **vaccines**, including dosage, adjuvantcy, route of immunization, and timing
- Qualifying and standardizing vaccine batches & protocols

## GENERAL INFORMATION

**Rabies** is a viral disease (Lyssavirus; ssRNA) that causes acute encephalitis (inflammation of the brain) in warm-blooded animals. It is transmitted by animals, most commonly by a bite from an infected animal, and is almost invariably fatal if post-exposure prophylaxis is not administered prior to the onset of severe symptoms.

Humans and animals have been protected, and the disease eradicated in certain geographical regions, by vaccination. Most vaccines have used whole inactivated virus that has been grown in a variety of cell types; vaccines using recombinant proteins of the rabies virus are also available. For rabies control of wildlife, vaccines in bait have proven effective. Improvement of the efficacy of vaccines is an active area of investigation.

The ADI anti-Rabies ELISA is designed with high sensitivity for discriminating lower level antibodies, with specially formulated diluents to minimize interfering background signals.

## PRINCIPLE OF THE TEST

The **Human Anti-Rabies IgG** ELISA kit is based on the binding of human anti- rabies in samples to rabies antigen immobilized on the microwells, and anti- rabies IgG antibody is detected by anti-human IgG-specific antibody conjugated to HRP (horseradish peroxidase) enzyme. After a washing step, chromogenic substrate (TMB) is added and color is developed by the enzymatic reaction of HRP on the substrate, which is directly proportional to the amount of anti-rabies IgG present in the sample. Stopping Solution is added to terminate the reaction, and absorbance at 450nm is then measured using an ELISA microwell reader. The activity of human antibody in samples is determined relative to anti-rabies calibrators.

## PRODUCT SPECIFICATIONS

### Specificity

Antigens prepared from whole-inactivated rabies virus subtypes 1-3 are used to coat the microwells; stabilizing postcoat contains BSA; thus, no other antibody specificity is detectable in the assay. The anti-Human IgG HRP conjugate primarily detects IgG, and does not react with IgM, IgA or IgE class antibodies above background.

### Assay Sensitivity

The rabies-coated plate and anti-Human IgG HRP concentration are optimized to differentiate anti-rabies IgG from background (non-antibody) signal with human serum samples diluted 1:100.

### Calibrator Values

The Calibrators are dilutions of anti-rabies antibody. Values are assigned as arbitrary anti-rabies virus activity units (see Limits of the Assay).

## KIT CONTENTS

The microtiter well plate and all other reagents, if unopened, are stable at 2-8°C until the expiration date printed on the box label. Stabilities of the working solutions are indicated under Reagent Preparation.

**To Be Reconstituted:** Store as indicated.

Component	Preparation Instructions
<b>Wash Solution Concentrate (100x)</b> Cat. No. WB-100, 10ml	Dilute the entire volume 10ml + 990ml with distilled or deionized water into a clean stock bottle. Label as <b>Working Wash Solution</b> and store refrigerated for long term and at lab temperature for short term.
<b>Sample Diluent Concentrate (20x)</b> Cat. No. SD-20T, 10ml	Dilute the entire volume, 10ml + 190ml with distilled or deionized water into a clean stock bottle. Label as <b>Working Sample Diluent</b> and store at 2-8°C until the kit lot expires or is used up.
<b>Anti-Human IgG-HRP Conjugate Concentrate (100x)</b> Part: H-HuG.2a11, 0.15ml	Peroxidase conjugated anti-human IgG in buffer with detergents and antimicrobial as stabilizers. Dilute fresh as needed; 10ul of concentrate to 1ml of <b>Working Sample Diluent</b> is sufficient for 1 8-well strip. Use within the working day and discard. Return 100X to 2-8°C storage.

**Ready For Use:** Store as indicated on labels.

Component	Part	Amt	Contents
<b>Rabies Antigen Coated Plate</b>	600-051	8-well strips (12)	Coated with rabies antigen, and post-coated with stabilizers.
<b>Anti-Rabies Calibrators</b>			
10 U/ml	600-042B	0.65 ml	Four (4) vials, each containing anti-rabies antibody in arbitrary activity Units; diluted in buffer with protein, detergents and antimicrobial as stabilizers.
25 U/ml	600-042C	0.65 ml	
50 U/ml	600-042D	0.65 ml	
100 U/ml	600-042E	0.65 ml	
<b>Anti-Rabies Positive Control</b>	600-042PC	0.65 ml	Serum with anti-rabies reactivity; [Value range on label]
<b>Low NSB Sample Diluent</b>	TBTm Not for HRP dilution	30 ml	Buffer with protein, detergents and antimicrobial. Use as is for sample dilution
<b>TMB Substrate</b>	80091	12 ml	Chromogenic substrate for HRP containing TMB and peroxide.
<b>Stop Solution</b>	80101	12 ml	Dilute sulfuric acid.

### Materials Required But Not Provided:

- Pipettors and pipettes that deliver 100ul and 1-10ml.
- Disposable glass or plastic 5-15ml tubes for diluting samples and Anti-Human IgG HRP Concentrate.
- Stock bottle to store diluted Wash Solution; 0.2 to 1L.
- Distilled or deionized water to dilute reagent concentrates.
- Microwell plate reader at 450 nm wavelength and ELISA plate washer

## LIMITATIONS OF THE ASSAY

### Calibrator Curve Quantitation

To quantitate antibody activity from a calibrator curve (such as provided with the kit), the dilution curve of the samples must be parallel to the calibrator curve, to avoid different values being obtained from different regions of the curve. Antibodies that are not matched in rabies avidity will often have non-parallel dilution curves. In these cases, antibody activity is best expressed as a titer relative to a reference positive such as the 25 U/ml Calibrator, or another Calibrator in the kit (see Calculation of Results).

## ASSAY DESIGN AND SET-UP

### Sample Collection and Handling

For **serum**, collect blood by venipuncture, allow clotting, and separate the serum by centrifugation at room temperature. For other samples, including **tissue culture media**, clarify the sample by centrifugation and/or filtration prior to dilution in Sample Diluent.

### Assay Design

Review Calculation of Results (p. 5-7) and Limits of the Assay (above) before proceeding:

- Select the proper sample dilutions accounting for expected potency of positives and minimizing non-specific binding (NSB) and other matrix effects; for example, net signal for non-immune samples should be <0.5 OD. This is usually 1:100 or greater dilution for human sera with normal levels of IgG and IgM.
- Run a Sample Diluent **Blank**. This signal is an indicator of proper assay performance, especially of washing efficacy, and is used for net OD calculations, if required (**See Method A, B**).
- Run a set of Calibrators. Calibrators validate that the assay was performed to specifications; results can be used to normalize between-assay variation for enhanced precision. Reading values off a Calibrator curve, **Method C**, has limitations. See Limits of the Assay (above).
- Run the **Anti-Rabies Positive Control**; value range is on the vial label..
- Run a range of sample dilutions for expected higher positives that allows calculation of antibody **Titer** when specific titer is at least 4-fold higher than non-immune). **See Method D**.
- Run samples in duplicate if used for quantitation; non-immunes that are significantly lower than immunes may be run in singlicate. The Calibrators that are used for quantitation, e.g., for between-assay normalization, should be run in duplicate. When determining titer from a dilution curve, singlicates can be run if more than two dilution points are used for titer calculations.

## Plate Set-up

Bring all reagents to room temperature (18-30° C) equilibration (at least 30 minutes).

- Determine the number of wells for the assay run. Duplicates are recommended, including 8 Calibrator wells and 2 wells for each sample and control to be assayed.
- Remove the appropriate number of microwell strips from the pouch and return unused strips to the pouch. Reseal the pouch and store refrigerated.
- Add 200-300ul Working Wash Solution to each well and let stand for about 1-5 minutes. Aspirate or dump the liquid and pat dry on a paper towel before sample addition.

## Assay Procedure

ALL STEPS ARE PERFORMED AT ROOM TEMPERATURE. After each reagent addition, gently tap the plate to mix the well contents prior to beginning incubation.

### 1. 1<sup>st</sup> Incubation [100ul – 60 min; 4 washes]

- Add 100ul of calibrators, samples and controls each to pre-determined wells.
- Tap the plate gently to mix reagents and incubate for 60 minutes.
- Wash wells 4 times and pat dry on fresh paper towels. As an alternative, an automatic plate washer may be used. Improper washes may lead to falsely elevated signals and poor reproducibility.

### 2. 2<sup>nd</sup> Incubation [100ul – 30 min; 5 washes]

- Add 100ul of diluted Anti-Human IgG HRP to each well.
- Incubate for 30 minutes.
- Wash wells 5 times as in step 1.

### 3. Substrate Incubation [100ul – 15 min]

- Add 100ul TMB Substrate to each well. The liquid in the wells will begin to turn blue.
- Incubate for 15 minutes in the dark, e.g., place in a drawer or closet.

Note: If your microplate reader does not register optical density (OD) above 2.0, incubate for less time, or read OD at 405-410 nm (results are valid).

### 4. Stop Step [Stop: 100ul]

- Add 100ul of Stop Solution to each well.
- Tap gently to mix. The enzyme reaction will stop; liquid in the wells will turn yellow.

### 5. Absorbance Reading

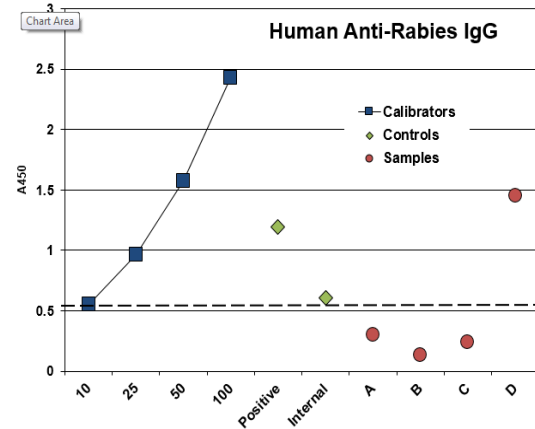
- Use any commercially available microplate reader capable of reading at 450nm wavelength. Use a program suitable for obtaining OD readings and data calculations if available.
- Read absorbance of the entire plate at 450nm using a single wavelength within 30 minutes after Stop Solution addition. If available, program to subtract OD at 630nm to normalize well background.

## INTERPRETATION OF RESULTS

### Method A. Antibody Activity Threshold Index

Compare Samples to **10 U/ml Calibrator** or **Internal Control**  
= **Positive/Negative Cut-off.**

#### Example:



#### Results

The **sensitivity** of the assay to detect anti-Rabies IgG, from either natural infection or vaccination, is controlled so that the **10 U/ml Calibrator** represents a threshold OD for most true positives in human serum diluted to 1:100 or greater. Visual inspection of the data in the above graph shows the following:

**Calibrators** – dilution curve of antiserum from rabies immunization, shows the OD range of the assay; high value indicates optimal sensitivity of the assay.

**10 U/ml:** a 'Cut-off' line has been drawn to indicate a threshold distinguishing between **Positive/Negative**. This is not a clear-cut threshold, rather a low OD area that could represent either low positives or high background negatives.

**Positive Control** – serum showing reactivity to rabies virus; the value range is on the label. This Control may be used to gauge precision and to normalize between-assay variation.

**Internal Control** – a true positive from an immune human that represents the investigator's experience in distinguishing low positive from negative samples (not in kit). This should be run in each assay to supplement the 1 U/ml Calibrator for Positive/Negative discrimination purposes.

**Samples A,B,C,D** – 3 samples (1:100) (A, B, C) are **negative**: below the threshold; 1 sample (D) is **positive**: clearly above the threshold.

The 10 U/ml Calibrator can be used to calculate a **Threshold Index** that numerically discriminates Positive/Negative:

- ❖ Divide each Sample net OD by the 10 U/ml Calibrator net OD. Values above 1.0 are a measure of **Positive** Antibody Activity; below 1.0 are **Negative** for antibody.

## INTERPRETATION OF RESULTS (cont)

### Method B. Positive Index

Experimental sample values may be expressed relative to the values of Control or Non-immune samples, by calculation of a **Positive Index**. One typical method is as follows:

1. Calculate the net OD mean + 2 SD of the Control/Non-immune samples = **Positive Index**.
2. Divide each sample net OD by the Positive Index. Values above 1.0 are a measure of **Positive** Antibody Activity; below 1.0 are **Negative** for antibody.

A sample value would be **Positive** if significantly above the value of the pre-immune serum sample or a suitably determined non-immune panel or pool of samples, tested at the same sample dilution.

This calculation also **quantifies** the positive Antibody Activity level, assigning a higher value to samples with higher Antibody Activity, and vice versa.

#### Example:

Sample	Assay Net OD		Calculated Antibody Activity	
	Control	Exptl	Control	Exptl
1	0.244	C 2.293	0.57	5.34
2	0.204	C 1.490	0.48	3.47
3	0.237	C 0.833	0.55	1.94
4	0.26	C 0.326	0.61	0.76
5	0.388	P 1.106	0.90	2.58
6	0.407	I 0.310	0.95	0.72
7	0.288	E 0.672	0.67	1.56
8	0.263	E 0.363	0.61	0.85
9	0.322	E 0.560	0.75	1.31
10	0.343	E 0.490	0.80	1.14
Mean	0.295			
SD	0.067			
Mean +2 SD	0.429	= <b>Positive Index</b>		

#### Results

**Experimental** Samples are represented as follows:

**C** – Calibrator  
**P** – Positive Control  
**I** – Internal Control; lab's threshold positive serum  
**E** – Experimental sample

## INTERPRETATION OF RESULTS (cont)

### Method C. Titers from Sample Dilution Curves

The titer of elevated antibody activity calculated from a dilution curve of each sample is recommended as the most accurate quantitative method. Best precision can be obtained using the following guidelines:

1. Use an OD value Index in the mid-range of the assay (2.0 – 0.5 OD); this provides the best sensitivity and reproducibility for comparing experimental groups and replicates. An arbitrary 1.0 OD is commonly used.
2. Prepare serial dilutions of each sample to provide a series that will produce signals higher and lower than the selected index. With accurate diluting, duplicates may not be required if at least 4 dilutions are run per sample.
3. A 5-fold dilution scheme is useful to efficiently cover a wide range which produces ODs both above and below 1.0 OD. The dilution scheme can be tightened to 3-fold or 2-fold for more precise comparative data.
4. The Calibrator values can be used to normalize inter-assay values.

#### Calculations

1. On a log scale of inverse of Sample Dilution as the x-axis, plot the OD values of the two dilutions of each positive sample having ODs above and below the OD value of the Index (arbitrary or selected Calibrator).
2. From a point-to-point line drawn between the two sample ODs, read the dilution value (x-axis) corresponding to the OD of the selected Index  
= **IgG Antibody Activity Units**

#### Notes:

- The sensitivity of the assay may be increased to perhaps convert a borderline sample to a positive by using a lower dilution of the sample, e.g., 1/50. The values of negatives may increase, so an alternative threshold should be established using known negatives to develop a **Positive Index** (page 6), or by using known **Internal Controls** as discriminator for a **Threshold Control** (instead of the kit 10 U/ml Calibrator Control)

#### Limits of the Assay

- Anti-rabies virus antibody levels of an infected or vaccinated host may be below detection threshold related to the time course of the occurrence, e.g., too early for positive titer development.
- To quantitate antibody activity from a calibrator curve (such as provided with the kit), the dilution curve of the samples must be parallel to the calibrator curve, to avoid different values being obtained from different regions of the curve. In cases of non-parallelism, antibody activity is best expressed as a titer relative to the titer of a reference positive, as shown above.

## PRECAUTIONS AND SAFETY INSTRUCTIONS

Calibrators, Sample Diluent, and Antibody HRP contain bromonitrodioxane (BND: 0.05%, w/v). Stop Solution contains dilute sulfuric acid. Follow good laboratory practices and avoid ingestion or contact of any reagent with skin, eyes, or mucous membranes. All reagents may be disposed of down a drain with copious amounts of water. MSDS for TMB, sulfuric acid, and BND can be requested or obtained from the ADI website.

# Human Anti-Rabies IgG ELISA Kit

Cat. # 600-020-HRV, 96 tests

For Quantitation of Anti-Rabies Virus IgG  
in serum, plasma or other biological  
fluids

For research use only (RUO), not for diagnosis, cure or  
prevention of the disease.



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ELISA Kit Components	Amount	Part
Rabies Coated Microwell Strip Plate	8-well strips (12)	600-051
Anti-Rabies Positive Control	0.65 ml	600-042PC
Anti-Rabies Calibrator 10 U/ml	0.65 ml	600-042B
Anti-Rabies Calibrator 25 U/ml	0.65 ml	600-042C
Anti-Rabies Calibrator 50 U/ml	0.65 ml	600-042D
Anti-Rabies Calibrator 100 U/ml	0.65 ml	600-042E
Anti-Human IgG HRP Conjugate (100X)	0.15 ml	H-HuG.2a11
Sample Diluent (20X)	10 ml	SD20T
Low NSB Sample Diluent	30 ml	TBTm
Wash Solution Concentrate (100X)	10 ml	WB-100
TMB Substrate	12 ml	80091
Stop Solution	12 ml	80101
Product Manual	1 ea	M-600-020-HRV