PERFORMANCE CHARACTERISTICS (continued)

Sample Recovery
High and low concentrations of mouse IgM were spiked into each of 3 serum samples. Observed assay values compared to expected values ranged from 87 to 96%, indicating accurate quantification of IgM in mouse serum.

Sample	Expected ng/ml	Observed ng/ml	Observed/ Expected
High IgM Spike		41.2	
+ Mouse D, 87.9 ng/ml	129.1	112.8	**87 %**
+ Mouse E, 152.3 ng/ml	193.5	182.8	**94 %**
+ Mouse F, 86.8 ng/ml	128.0	123.0	**96 %**
Low IgM Spike		11.2	
+ Mouse D, 87.9 ng/ml	99.1	89.3	**90 %**
+ Mouse E, 152.3 ng/ml	163.5	144.5	**88 %**
+ Mouse F, 86.8 ng/ml	98.0	92.0	**94 %**

Related Items

Catalog# ProdDescription
6310 Mouse IgA ELISA Kit, 96 tests, Quantitative
6320 Mouse IgG ELISA Kit, 96 tests, Quantitative
6330 Mouse IgG1 ELISA Kit, 96 tests, Quantitative
6340 Mouse IgG2a ELISA Kit, 96 tests, Quantitative
6350 Mouse IgG2b ELISA Kit, 96 tests, Quantitative
6360 Mouse IgG3 ELISA Kit, 96 tests, Quantitative
6370 Mouse IgE ELISA Kit, 96 tests, Quantitative
6380 Mouse IgM ELISA Kit, 96 tests, Quantitative
6380-RS Mouse IgM Reference Serum for ELISA (~500 ng/ml)
6390 Mouse Transferrin (Tf) ELISA Kit, 96 tests, Quantitative

6320-RDT-25 TruStrip RDT Mouse IgG Rapid Test cards, 10/pk

For more details please consult our web site (www.4adi.com) or contact us by email (service@4adi.com).

Instruction Manual No. M-6380

Mouse IgM ELISA Kit

Cat. No. 6380, 96 Tests

For Quantitative Determination of Mouse
Immunoglobulin M in serum, plasma or other biological Fluids

For research use only (RUO), not for diagnosis, cure or prevention of the disease.

ALPHA DIAGNOSTIC INTERNATIONAL

6203 Woodlake Center Drive • San Antonio• Texas 78244 • USA.
Phone (210) 561-9515 • Fax (210) 561-9544
Toll Free (800) 786-5777

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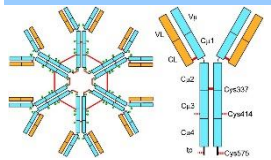
www.4adi.com

INTENDED USE

The Mouse IgM ELISA Kit is an in vitro immunoassay for the quantification of IgM circulating in serum or in other appropriately qualified samples from tissue fluids (e.g., saliva, mucosa), or in cultures of mouse cells. For research use only (RUO), not for diagnosis, cure or prevention of the disease.

RESEARCH USE OF THE TEST

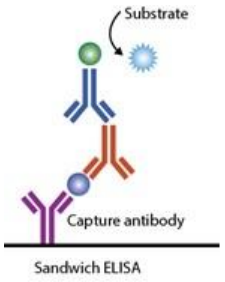


Immunoglobulin M (IgM) is a basic antibody that is produced by B cells. IgM is by far the physically largest antibody in the circulatory system. It is the first antibody to appear in response to initial exposure to an antigen. IgM is produced and secreted from spleen, a major site for B cell activity. IgM forms polymers where multiple immunoglobulins are covalently linked together with disulfide bonds, mostly as a pentamer (970 kda) but also as a hexamer. Because each monomer has two antigen binding sites, a pentameric IgM has 10 binding sites. The J chain is found in pentameric IgM but not in the hexameric form. Due to its polymeric nature, IgM possesses high avidity, and is particularly effective at complement activation. It contributes greatly to opsonization by activating complement and causing C3b to bind to the antigen. IgM normal human plasma concentration ~1.-4 mg/ml. Low levels of IgM are associated with Wiskott-Aldrich syndrome.

Immunoassays using heavy-chain specific antibodies provide for selective, sensitive quantification of mouse immunoglobulins IgG, IgA and IgM, as found circulating in blood or as present in other body fluids, including saliva, milk/colostrum, ascites, tears and mucosa of linings of the gut, respiratory or urigenital tracts. Levels of total IgG, IgA and/or IgM can reveal health status or results of experimental or pathological conditions (e.g., hypo- or hypergammaglobulinemia or acute or chronic infection). Also, measurements of specific antibody levels, in antigen-specific assays, are often best interpreted relative to values of total IgM, IgA, and IgM in the sample and/or individual.

The assays are also suitable for quantifying monoclonal antibodies and can be a reliable method for monitoring production and establishing quality control. The quantitative immunoassays measure mouse IgG, IgA and IgM with high sensitivity; this allows dilution beyond interference from the sample matrix for samples derived from any of the above specimen types. Also, each assay is Ig class specific, such that all IgG or IgA subclasses are reliably quantified in essentially any specimen, freshly obtained and/or suitable stored. Expected performance of each kit relative to precision, recovery and linearity of dilution is presented as guidance for use and experimental design.

PRINCIPLE OF THE TEST



The Mouse IgM ELISA kit is based on the binding of mouse IgM in samples to two antibodies, one immobilized on the microtiter wells, and the other conjugated to horseradish peroxidase (HRP) enzyme. After a washing step, chromogenic substrate is added and color is developed by the enzymatic reaction of HRP on the TMB substrate, which is directly proportional to the amount of IgM present in the sample. Stopping Solution is added to terminate the reaction, and absorbance at 450nm is then measured using an ELISA microtiter well reader. The concentration of IgM in samples and control is calculated from a curve of standards containing known concentrations of IgM.

PERFORMANCE CHARACTERISTICS

Specificity

The antibodies used in this kit have been shown by immunoelectrophoresis and ELISA to react specifically with IgM, and have essentially no reactivity with IgG, IgA, IgE or any other mouse serum proteins.

Serum from the following species showed no significant reactivity at 1:400 dilution: human, rat, hamster, guinea pig, bovine, pig, horse, sheep, goat, dog, cat, rabbit or chicken; also 10% neonatal bovine serum.

Normal Range

Assay of IgM in stored sera from twenty (20) individual Swiss mice ranged from 22 to 712 ug/ml (median = 519 ug/ml). Each laboratory should determine expected values of its own testing population.

Precision

Samples containing low, medium and high concentrations of IgM, representing 3 different sera, were assayed multiple times in the same assay (n=10) to provide within-assay precision, and as duplicates in multiple assays (n=5) to obtain between-assay reproducibility. Coefficient of variations were calculated for the concentrations using a point-to-point curve-fitting program.

IgM concentrations were measured with very good within-assay (3.9 to 6.7 %CV) and between-assay (5.2 to 8.5 %CV) reproducibility.

Sample	IgM ng/ml	Intra-assay %CV	Inter-assay %CV
Mouse A	30	**6.7**	**5.2**
Mouse B	72	**3.9**	**5.7**
Mouse C	108	**5.7**	**8.5**

Linearity of Dilution

Three (3) individual pooled stored sera and one (1) monoclonal ascites were diluted to 2 levels for testing, and concordance of the assay values were compared. The mean recovery ranged from 95 to 100%, demonstrating linear dilution and equivalent quantification across the standard range.

Sample	Dilution	Assay Value ng/ml	Serum Value ug/ml	Concordance
Mouse Pool A	1:800 1:6400	102 12	82 77	**97 %**
Mouse Pool B	1:800 1:6400	176 22	141 141	**100 %**
Mouse Pool C	1:800 1:6400	111 15	89 99	**95 %**
Mouse Ascites	1:5k 1:20k	33.8 7.9	169 157	**96 %**

CALCULATION OF RESULTS

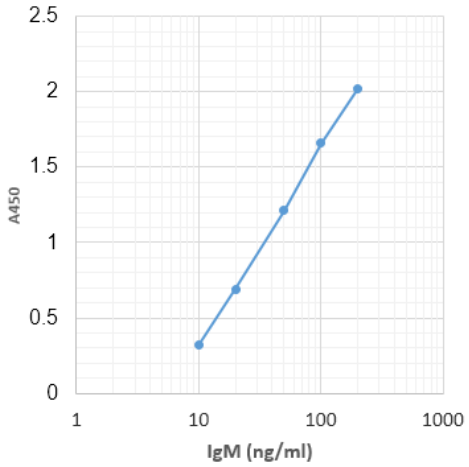
1. The results may be calculated using any immunoassay software package. The four-parameter curve-fit is recommended. If software is not available, IgM concentrations may be determined as follows:
2. Calculate the mean OD of duplicate samples.
3. On graph paper plot the mean OD of the standards (y-axis) against the concentration (ng/ml) of IgM (x-axis). Draw the best fit curve through these points to construct the standard curve. A point-to-point construction is most common and reliable.
4. The IgM concentrations in unknown samples and controls can be determined by interpolation from the standard curve.
5. Multiply the values obtained for the samples by the dilution factor of each sample.
6. Samples producing signals higher than the 200 ng/ml standard should be further diluted and re-assayed.

TYPICAL RESULTS

The following data are for illustration purposes only. A complete standard curve should be run in every assay to determine sample values.

Wells	Standards, Control & Samples	A450 nm	**IgM** ng/ml
A1, A2	**Negative Diluent Control**	0.03	0
B1, B2	10 ng/ml **Standard**	0.32	10
C1, C2	25 ng/ml **Standard**	0.69	25
D1, D2	50 ng/ml **Standard**	1.11	50
E1, E2	100 ng/ml **Standard**	1.66	100
F1, F2	200 ng/ml **Standard**	2.02	200
G1, G2	**Positive Serum Control** [Value: 88 -156 ng/ml]	1.67	103
H1, H2	**Sample** [Diluted 1:1600]	1.07	46
	Calculated: 1600-fold dilution x 46 ng/ml = **73.6** ug/ml in serum		

A typical assay Standard Curve (do not use for calculating sample values)



/bill/3-ELISA/6380

STORAGE AND STABILITY

The microtiter well plate and all other reagents, if unopened, are stable at 2-8^{o}C until the expiration date printed on the label. Stabilities of the working solutions are indicated under Reagent Preparation.

KIT CONTENTS

To Be Reconstituted: Store as indicated.

Component	Instructions for Use
Sample Diluent Concentrate (20x) Cat. No. SD-20T, 10ml	Dilute the entire volume, 10ml + 190ml with distilled or deionized water into a clean stock bottle. Label as **Working Sample Diluent** and store at 2-8ºC until the kit lot expires or is used up.
Wash Solution Concentrate (100x) Cat. No. WB-100, 10ml	Dilute the entire volume 10ml + 990ml with distilled or deionized water into a clean stock bottle. Label as **Working Wash Solution** and store at RT until kit is used entirely.
Anti-Mouse IgM - HRP Conjugate Concentrate (100x) Part No. 6384, 0.15ml	Peroxidase conjugated anti-mouse IgM in buffer with protein, detergents and ProClin 300 as stabilizers. Dilute fresh as needed; 10ul of concentrate to 1ml of **Working Sample Diluent** is sufficient for 1 8-well strip. Use within the working day and discard. Return concentrate to 2-8ºC storage.

Ready For Use: Store as indicated on labels.

Component	Part No.	Amt	Contents
Anti-Mouse IgM Microwell Strip Plate	6381	8-well strips (12)	Coated with purified anti-Mouse IgM antibodies.
Mouse IgM Standards			
10 ng/ml 25 ng/ml 50 ng/ml 100 ng/ml 200 ng/ml	6383B 6383C 6383D 6383E 6383F	0.65 ml 0.65 ml 0.65 ml 0.65 ml 0.65 ml	Five (5) vials, each containing mouse serum with calibrated IgM concentrations; diluted in buffer with protein, detergents and ProClin 300 as stabilizers.
Positive Control [IgM] range on label	6382	0.65 ml	Mouse serum with stated IgM concentration range; diluted in buffer with protein, detergents and ProClin 300 as stabilizers.
TMB Substrate	80091	12 ml	Chromogenic substrate for HRP containing TMB and peroxide.
Stop Solution	80101	12 ml	1% sulfuric acid.

Materials Required But Not Provided:

- Pipettors and pipettes that deliver 100ul and 1-10ml. A multi-channel pipetter is recommended.
- Disposable glass or plastic 5-15ml tubes for diluting samples and Anti-mouse IgM-HRP Concentrate.
- Graduated cylinder to dilute Wash Concentrate and Sample Diluent concentrate; 200ml to 1L.
- Stock bottle to store diluted Wash Solution; 200ml to 1L.
- Distilled or deionized water to dilute reagent concentrates.
- Microwell plate reader at 450 nm wavelength.

SPECIMEN COLLECTION AND HANDLING

Collect blood by venipuncture, allow clotting, and separate the serum by centrifugation at room temperature. Do not heat inactivate the serum. If sera are not assayed immediately , stored refrigerated for up to 2 weeks, or frozen for long-term storage. Avoid freeze-thaw cycles.
The use of plasma has not been investigated, but should be a suitable specimen for assay.

PRECAUTIONS AND SAFETY INSTRUCTIONS

Standards, Controls, Sample Diluent, and Anti-mouse IgM-HRP contain Proclin 300 (0.05%, v/v). Stop Solution contains 1% sulfuric acid. Follow good laboratory practices, and avoid ingestion or contact of any reagent with skin, eyes or mucous membranes. All reagents may be disposed of down a drain with copious amounts of water.

Applicable MSDS, if not already on file, for the following reagents can be obtained from ADI or the web site for Prolcin-300 (0.1% v/v in standards, and assay buffers).
http://4adi.com/commerce/info/showpage.jsp?page_id=1060&category_id=2430&visit=10

QUALITY CONTROL

Reagents Accurate and reproducible assay results rely on proper storage, handling and control of reagent and sample temperature. Store all reagents as indicated, and warm to room temperature only those to be used in the assay. Shelf-life of the critical reagents and samples will diminish with extended exposure to non-refrigeration, resulting in inaccurate assay results. All solutions should be clear. Cloudiness or particulates are indications of reagent contamination or instability and may interfere with proper performance of the assay. Do not use.

Sample Controls A Positive Serum Control is provided with the kit, assigned with an IgM concentration value range. Recovery in this range is an indicator of proper assay performance. Each lab should also assay internal control samples, which represent the lab's expected sample population and that are maintained stabilized. A Negative Diluent Control should also be run.

Standard Curve The signal generated by the standards should be continuously increasing in OD from the lowest Standard to the highest Standard, with a difference greater than 1.2 OD. Non-continuously increasing or low signals may indicate problems with technique, protocol directions and/or reagent preparation, use or stability. A Negative Diluent Control should be of lower signal than the lowest standard. Do not rely on results generated from an assay with these issues.

Technique Accurate and reproducible assay results rely on good lab technique regarding pipetting, plate washing and handling of samples and reagents.

Equipment Precision of results relies on uniform and effective washing techniques; an automatic washer may be used. ELISA reader and pipettes should be properly calibrated.

ASSAY PROCEDURE

Bring all reagents to room temperature (18-30° C) equilibration (at least 30 minutes).

DILUTE Serum Samples in Working Sample Diluent. Dilutions of about 1:1-4k are appropriate for most normal mouse sera. For accuracy, two dilution steps are recommended, as follows:
1) 10ul serum + 490ul diluent = [1:50],
2) 10ul [1:50] + 390ul diluent = **[1:2000]**.

DO NOT dilute the Standards or Positive Control Serum.

ALL STEPS ARE PERFORMED AT ROOM TEMPERATURE. After each reagent addition, gently tap the plate to mix the well contents prior to beginning incubation.

1. Set-up

- Determine the number of wells for the assay run. Duplicates are recommended, including 10 Standard wells and 2 wells for each sample and control to be assayed.
- Remove the appropriate number of microwell strips from the pouch and return unused strips to the pouch. Reseal the pouch and store refrigerated.
- Add 200-300ul Working Wash Solution to each well and let stand for 5 to 30 minutes before sample addition.
- Aspirate the liquid and pat dry on a paper towel.

2. 1st Incubation [100ul - 60min; 4 washes]

- Add 100ul of standards, samples and controls each to pre-determined wells.
- Tap the plate gently to mix reagents and incubate for 60 minutes.
- Wash wells 4 times and pat dry on fresh paper towels. As an alternative, an automatic plate washer may be used. Improper washes may lead to falsely elevated signals and poor reproducibility.

3. 2nd Incubation [100ul - 30min; 5 washes]

- Add 100ul of diluted Anti-mouse IgM-HRP Conjugate to each well.
- Incubate for 30 minutes.
- Wash wells 5 times as in step 2.

4. Substrate Incubation [100ul - 15min]

- Add 100ul TMB Substrate to each well. The liquid in the wells will begin to turn blue.
- Incubate for 15 minutes in the dark, e.g., place in a drawer or closet.

Note: If your microplate reader does not register optical density (OD) above 2.0, incubate for less time, or read OD at 405-410 nm (results are valid).

5. Stop Step [Stop: 100ul]

- Add 100ul of Stop Solution to each well.
- Tap gently to mix. The enzyme reaction will stop; liquid in the wells will turn yellow.

6. Absorbance Reading

- Use any commercially available microplate reader capable of reading at 450nm wavelength. Use a program suitable for obtaining OD readings, and data calculations if available.
- Read absorbance of the entire plate at 450nm using a single wavelength within 30 minutes after Stop Solution addition. If available, program to subtract OD at 630nm to normalize well background.